ORIGINAL ARTICLE

Open Access

The specific status of *Melastoma kudoi* (Melastomataceae, Melastomeae)



Jin-Hong Dai¹, Che-Wei Lin², Qiu-Jie Zhou¹, Chun-Mei Li¹, Ren-Chao Zhou^{1*} and Ying Liu^{1*}

Abstract

Background: *Melastoma* has undergone rapid species radiation during the last one million years, and circumscription of some species in the genus has remained controversial. *Melastoma kudoi*, an erect species narrowly endemic to central Taiwan was previously treated as a synonym of *M. intermedium*, a semicreeping hybrid between the erect species *M. candidum* and the creeping *M. dodecandrum*, making its identity questionable. We addressed this question based on molecular and morphological data.

Results: Phylogenetic analyses based on nrITS sequence data revealed that *M. kudoi* is most closely related to *M. dodecandrum*. Further analyses of six nuclear genes (*cam, chi, gapC, gbss, tpi* and *vr*) and two chloroplast markers (*trnL–trnF* and *psbA*) showed that *M. kudoi* is well diverged from its close relatives. Morphologically, it is also easily distinguished from related species by its erect habit, center-positioned stigma, and spreading, basally enlarged hairs on the hypanthium.

Conclusions: Both molecular phylogenetic and morphological data suggest that *M. kudoi* is well separated from *M. intermedium*, *M. dodecandrum*, and *O. scaberrima*, and should be treated as a distinct species. Taxonomic treatment and detailed description of *M. kudoi* are provided.

Keywords: Melastoma kudoi, Otanthera, Phylogeny, Morphology, Taxonomy, Taiwan

Background

Melastoma L. is distributed in tropical Asia and Oceania (Meyer 2001; Chen and Renner 2007). Previous study indicated that this genus has undergone rapid species radiation during the last one million years (Renner and Meyer 2001), and natural hybridization is commonly found among species with overlapping geographical range and flowering time (Dai et al. 2012; Liu et al. 2014; Zhou et al. 2017; Zou et al. 2017). The exact number of species in *Melastoma* is unclear. Description of new species from poorly surveyed areas would be expected as many are narrowly endemic (Wong 2016; Neo et al. 2017).

Generic circumscription of *Melastoma* has remained controversial. Some authors recognized the Asian genus

Otanthera Blume as distinct from Melastoma (e.g. Keng and Li 1977; Chen 1984; Huang and Huang 1993), while some merged it into *Melastoma* (Meyer 2001; Yang and Liu 2002; Chen and Renner 2007). Species delimitation in Melastoma is also problematic. Multiple names have often been applied for the same taxa across different works (Keng and Li 1977; Chen 1984; Huang and Huang 1993; Meyer 2001; Yang and Liu 2002; Chen and Renner 2007; Dai et al. 2012; Chao et al. 2014; Huang et al. 2018). Regretfully, these issues have not been fully addressed using an integrative approach. In this study, we basically followed the taxonomy of Chen (1984) for M. candidum D. Don, M. dodecandrum Lour., M. normale D. Don, M. sanguineum Sims, and Otanthera scaberrima (Hayata) Ohwi. For M. malabathricum L., which was not included in Chen (1984), we adopted a circumscription modified from Meyer (2001).

Melastoma kudoi Sasaki was initially described based on a collection, Kudo & Sasaki s.n. (Fig. 1a), from Lake Sui-sya (Sun Moon Lake), Taiwan (Sasaki 1931). Keng



^{*}Correspondence: zhrench@mail.sysu.edu.cn; liuyng73@mail.sysu.edu.cn

State Key Laboratory of Biocontrol and Guangdong Key Laboratory
of Plant Resources, School of Life Sciences, Sun Yat-sen University,
Guangzhou 510275, China

Full list of author information is available at the end of the article

Dai et al. Bot Stud (2019) 60:5 Page 2 of 11



Fig. 1 a Taiwan, Lake Sui-sya, Kudo & Sasaki s.n. (TAI), type of Melastoma kudoi Sasaki, image from TAI Herbarium; b China, Fujian, Yenping, Dunn 2706 (A), isotype of Melastoma intermedium, image from JSTOR

and Li (1977) didn't recognize *M. kudoi* as a distinct species, merging it within *M. intermedium* Dunn. This treatment was followed by Chen (1984) and Huang and Huang (1993). Meyer (2001) synonymized *M. intermedium* under *M. malabathricum*, and listed *M. kudoi* as species dubiae in his revision of Southeast Asian *Melastoma*, because too little information about the latter species was available. Yang and Liu (2002) noted that the plants previously identified as *M. intermedium* in Taiwan were different from *M. malabathricum*, they therefore applied the earliest published name *M. kudoi* for these plants. Chen and Renner (2007), in the most recent revision of Chinese species of *Melastoma*, placed *M. kudoi* under *M. intermedium* together with *M. suffruticosum* Merr. and *Otanthera scaberrima*.

As shown in Fig. 1b, the isotype of *Melastoma intermedium* (Dunn 2706) is a semi-creeping plant with its lower part of stem growing horizontally and the upper part vertically. This species is intermediate between the erect *M. candidum* and *M. dodecandrum*, the only creeping species in *Melastoma*, in terms of habit, bract

size and indumentum on the hypanthium (Fig. 2), and it is often observed sympatrically with the latter two species in southeastern mainland China (Dai et al. 2012). In accordance with morphology, DNA sequence data had revealed that *M. intermedium* is a natural hybrid between M. candidum and M. dodecandrum (Dai et al. 2012). Although most authors treated M. kudoi as a synonym of M. intermedium, both herbarium specimen (Fig. 1a) and type protologue (Sasaki 1931) indicate that M. kudoi is in fact an erect shrub quite different from the semi-creeping M. intermedium (Fig. 1b). Geographically, M. kudoi is narrowly endemic to central Taiwan, where M. dodecandrum, one parental species of M. intermedium, has never been recorded. Thus, its habit plus geographical range suggest that M. kudoi may represent a distinct species rather than a conspecific of *M. intermedium*.

In this paper, we try to evaluate the specific status of *M. kudoi* based on molecular and morphological data. First, a phylogenetic hypothesis of *Melastoma* was reconstructed using sequence data of nrITS to determine the phylogenetic position of *M. kudoi* in the genus. Six

Dai et al. Bot Stud (2019) 60:5 Page 3 of 11



Fig. 2 *Melastoma intermedium* Dunn. **a** Habit; **b** flowering branches; **c** adaxial and abaxial surface of leaf; **d** bracts on the inflorescence; **e** flower, arrow indicated the position of stigma; **f** young fruits showing the appressed hairs

low-copy nuclear genes (cam, chi, gapC, gbss, tpi and vr) and two chloroplast markers (trnL-trnF and psbA) were then sequenced and analyzed for selected outgroup and ingroup taxa to test the distinctness of M. kudoi. Morphological comparison is made among M. kudoi and its closest relative as revealed by phylogenetic analyses.

Methods Sampling

We sampled three individuals per population from one population of *M. candidum* (Longhai, Fujian), one of *M. kudoi* (Nantou, Taiwan), one of *Otanthera scaberrima* (Pingdong, Taiwan), and three of *M. dodecandrum*

Dai et al. Bot Stud (2019) 60:5 Page 4 of 11

(Pinghe, Fujian; Ruyuan, Guangdong; Cangnan, Zhejiang). For each sample, fresh leaves were collected in the field and dried with silica gel until DNA extraction. Aside from the above taxa, four other species of *Melastoma* and one species of *Osbeckia L.*, viz. *M. intermedium*, *M. malabathricum*, *M. normale*, *M. sanguineum*, and *Osbeckia nepalensis* Hook. were also combined into the analyses using nrITS sequences downloaded from GenBank. A complete list of the taxa sampled in this study, their collection localities, voucher information, and GenBank accession numbers are provided in Additional file 1: Table S1.

DNA extraction, PCR amplification and sequencing

Total DNA was extracted from dried leaf tissue using the modified CTAB procedure (Doyle and Doyle 1987). Six low-copy nuclear genes (cam, chi, gapC, gbss, tpi and vr) were amplified and sequenced using primers published in previous studies of *Melastoma* (Dai et al. 2012; Chao et al. 2014; Zhou et al. 2017; Huang et al. 2018). For nrITS and chloroplast intergenic spacer trnL-trnF, universal primers were used (White et al. 1990; Taberlet et al. 1991). In addition, we also amplified and sequenced partial chloroplast gene psbA using primers (psbA-F: 5'-TACGCAACA GCAATCCAAGG-3', psbA-R: 5'-AGATATTGGTTG ACACGGGGA-3') designed based on the chloroplast genome sequence of M. candidum (Ng et al. 2017). PCR amplification and sequencing were conducted following the same experimental procedure described in Zou et al. (2017). For some samples with multiple polymorphic sites, cloning sequencing was used to separate the haplotypes. Clonings were performed with the pMD18-T and A cloning kit (Takara, Dalian, China) and six positive colonies were sequenced. A total of 162 sequences were newly generated for phylogenetic analyses (Additional file 1: Table S1), whereas 6 were downloaded from GenBank.

Molecular analysis

The nrITS sequences were assembled and aligned in Seq-Man 7.1.0 (DNASTAR Inc., Madison, WI). Osbeckia nepalensis was adopted as an outgroup according to recent study of Melastomataceae (Veranso-Libalah et al. 2017). The nrITS dataset included two haplotypes of the hybrid taxon M. intermedium, one identical to the sequences of M. candidum, M. malabathricum, M. normale and M. sanguineum, and the other identical to those of M. dodecandrum and M. kudoi. Therefore, only one sequence each from the two clusters were used in the analysis. A maximum likelihood tree was constructed with RAxML 8.2.9 (Stamatakis 2014) using default parameters to explore the phylogenetic affiliation of M. kudoi in the

genus. Based on the resulted phylogeny, outgroup and ingroups were selected for subsequent analyses.

Sequences of six nuclear genes were concatenated and aligned using MAFFT 7.307 (Katoh and Standley 2013). For the species with no intraspecific sequence variation, the sequence of one individual was used in the analyses. The best-fitting model for each gene was determined using Modeltest 3.7 (Posada and Crandall 1998) with the Akaike information criterion (AIC). The substitution models GTR, K81uf, TrN, TrN, HKY and F81 were selected for the cam, chi, gapC, gbss, tpi and vr regions respectively. Bayesian inference (BI) analyses were conducted in MrBayes 3.2 (Ronquist et al. 2012). The Markov Chain Monte Carlo (MCMC) analysis was conducted with four simultaneous chains of 1,000,000 generations, sampling one tree every 100 generations. We verified that the average deviation of split frequencies had reached a value below 0.01. The first 2500 trees (25%) were discarded as burn-in, and the remaining trees were used to construct a 50% majority-rule consensus tree with Bayesian posterior probabilities. Maximum parsimony (MP) analyses were performed in PAUP* version 4.0b10 (Swofford 2002). A heuristic search strategy was carried out of 1000 random addition replicates, with tree-bisection-reconnection (TBR) branch swapping algorithm and MultTrees on. Maxtree was set to 500. The trees were evaluated by 1000 bootstrap replicates of 1000 random additions. Maximum Likelihood (ML) analyses were performed in RAxML 8.2.9 (Stamatakis 2014) with default parameters. The interspecific genetic distances were also calculated with Kimura 2-Parameter model (Kimura 1980) using MEGA 7 (Kumar et al. 2016).

Sequences of chloroplast *trnL-trnF* and *psbA* regions were concatenated and used to construct a haplotype network diagram in Network 5.0.0.3 (http://www.fluxus-engineering.com/) with the median-joining algorithm (Bandelt et al. 1999).

Morphological observation

For observation of the habit and other morphological characters of *M. kudoi*, we visited its type locality and transplanted two individuals into the greenhouse of Taiwan Forestry Research Institute in October, 2016. The living individuals were kept till its flowering stage. The habit, leaves, inflorescence, and detail of the flower were then recorded using a digital camera and compared with its close relatives.

Results

Molecular analysis

The nrITS dataset contained 653 characters. Phylogenetic analysis revealed two sister clades: one included *M. candidum, M. malabathricum, M. normale, M.*

Dai et al. Bot Stud (2019) 60:5 Page 5 of 11

sanguineum, and haplotype 1 of *M. intermedium*, and the other contained *M. dodecandrum*, *M. kudoi*, haplotype 2 of *M. intermedium*, and *Otanthera scaberrima* (Fig. 3). Based on these results, *M. candidum* was selected as an outgroup, and *M. kudoi*, *M. dodecandrum* and *Otanthera scaberrima* as ingroups for subsequent analyses.

The concatenated matrix of the six nuclear genes were 4322 bp long. No intraspecific sequence variation was observed for *M. kudoi* and *Otanthera scaberrima*.

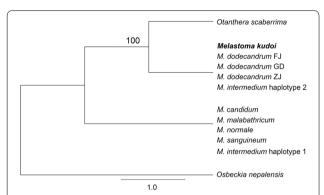


Fig. 3 Maximum likelihood (ML) phylogenetic tree based on the nrITS dataset. Number above branch is bootstrap value obtained from maximum likelihood analysis. FJ, GD and ZJ denote the three sampling localities of *M. dodecandrum*. For the species with identical ITS sequences, only one sequence was used in the analysis

The tree topologies constructed using ML, BI and MP algorithms were identical. Here we presented only the ML tree, with Bayesian posterior probabilities and MP bootstrap support values also marked on the branches. As shown in Fig. 4, individuals of *M. dodecandrum*, *M. kudoi* and *Otanthera scaberrima* clustered in separate subclades of their own; the three subclades then formed a well-recognized clade (PP=1, BS ML=100%, BS MP=100%), within which *O. scaberrima* was the early diverged branch and *M. kudoi* was sister to *M. dodecandrum* with strong support (PP=1, BS ML=93%, BS MP=93%). As shown in Table 1, molecular divergence between *M. kudoi* and the rest species, viz. *M. dodecandrum*, *M. candidum* and *O. scaberrima*, were 0.00609, 0.01387 and 0.01665 respectively.

The total length of the chloroplast dataset was 1292 bp, no intraspecific variation was detected in each of the four species. There was one differentially fixed nucleotide substitution between *M. kudoi* and *M. dodecandrum*. Two to four mutational steps were observed among other species (Fig. 5).

Morphology

Field observation showed that the only population of *M. kudoi* currently known grew at the base of a dam by Lake Sui-sya where plant individuals were sometimes mowed because of intense human maintenance (Fig. 6).

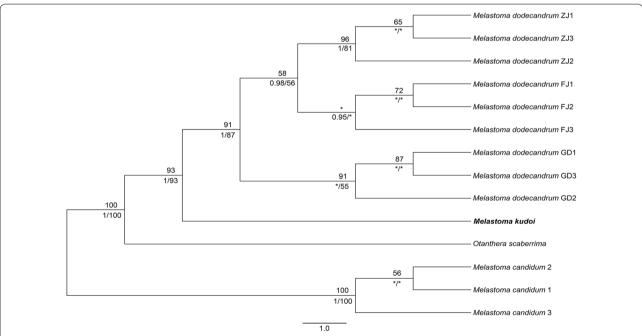


Fig. 4 Maximum likelihood (ML) phylogenetic tree based on the concatenated 6 nuclear genes. Numbers above branches are bootstrap values obtained from ML analysis, and those below branches are Bayesian posterior probabilities (left) and bootstrap values (right) resulting from MP analysis. FJ, GD and ZJ denote the three sampling localities of *M. dodecandrum*. For species with no intraspecific sequence variation, the sequences of one individual was used in the analyses. Asterisks indicate low support values (PP < 0.90, BS < 50%)

Dai et al. Bot Stud (2019) 60:5 Page 6 of 11

Table 1 Pairwise genetic distances among M. kudoi and three closely related species based on six nuclear genes

	Melastoma kudoi	Melastoma dodecandrum	Melastoma candidum	Otanthera scaberrima
Melastoma kudoi				
Melastoma dodecandrum	0.00609			
Melastoma candidum	0.01387	0.00823		
Otanthera scaberrima	0.01665	0.01047	0.01715	

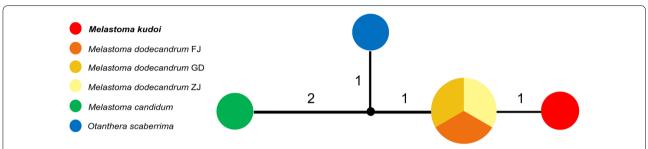


Fig. 5 Haplotype network of the combined chloroplast regions for the 6 populations. Haplotypes of each population are indicated by different colors. The numbers on the connecting lines denote the mutational steps. The black dot represents hypothesized haplotype. FJ, GD and ZJ denote the three sampling localities of *M. dodecandrum*



Fig. 6 Melastoma kudoi Sasaki. a Habitat; b plant individuals under intense human maintenance

One of the two transplanted individuals entered its flowering stage in June, 2017. The mature individual of $M.\ kudoi$ is an erect shrub (Fig. 7a) ca. 40 cm tall. The leaves are opposite, ca. 4×2 cm, oval or oblong oval (Fig. 7b), remotely strigose, petiole 2 mm long. Terminal cymes are composed of 1–3 flowers (Fig. 7c). Flowers are about 5.5 cm in diameter, bearing 10 stamens dimorphic in both length and morphology (Fig. 7d). The style and stigma are positioned in the center of the flower amongst the shorter stamens (Fig. 7e, f). Hypanthium and young fruit are about 9×7 mm in size, and densely strigose with spreading, basally enlarged hairs (Fig. 7g).

Discussion

Molecular evidence supported the specific status of *M. kudoi*. Apart from the hybrid taxon *M. intermedium*, phylogenetic analyses based on nrITS sequence data revealed that *M. kudoi* is most closely related to *M. dodecandrum* and *Otanthera scaberrima* (Fig. 3). Further analyses based on sequence data of six nuclear genes and two chloroplast markers confirmed the close relationship among these species (Figs. 4, 5). Genetic distances calculated from sequence data of six nuclear genes showed that molecular divergence between *M. kudoi* and other species (0.00609–0.01665) were comparable with those among other species (0.00823–0.01715) (Table 1),

Dai et al. Bot Stud (2019) 60:5 Page 7 of 11

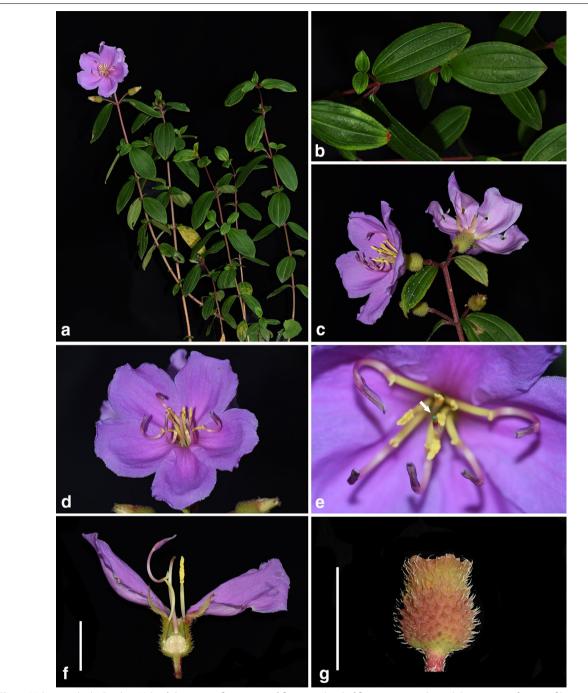


Fig. 7 *Melastoma kudoi* Sasaki. **a** Habit; **b** leaves; **c** inflorescence; **d** flower; **e** detail of flower, arrow indicated the position of stigma; **f** longitudinal section of a flower, scale bar 10 mm; **g** young fruit showing the dense, spreading, basally enlarged hairs, scale bar 10 mm

indicating that it is well diverged from *M. dodecandrum* and *Otanthera scaberrima*.

Melastoma kudoi is morphologically distinct from related species, viz. *M. intermedium*, *M. dodecandrum*, and *O. scaberrima*. Most previous authors had treated *M. kudoi* under *M. intermedium* based on gross morphology. This is quite understandable, as both species have leaves

and flowers of similar size. In addition, the semi-creeping branches of *M. intermedium* looked like they were erect when pressed and oriented on the herbarium sheet. Nevertheless, observation of living materials revealed that *M. kudoi* differs from *M. intermedium* in its wholly erect habit (vs. semi-creeping), center-positioned stigma near the shorter stamens (vs. side-positioned, near the

Dai et al. Bot Stud (2019) 60:5 Page 8 of 11

longer stamens), and its hypanthium densely strigose with spreading, basally enlarged hairs (vs. appressed hairs without enlarged bases) (Figs. 2, 7). Sequence data from both nuclear and chloroplast markers had placed *M. dodecandrum* as the closest relative of *M. kudoi*. However, the two species can be rather easily distinguished from each other by a series of characters such as habit

(erect vs. creeping), flower size (5.5 cm vs. 2.5–4 cm in diameter), hypanthium size (9 mm vs. 5 mm long), and stigma position (center-positioned vs. side-positioned) (Figs. 7, 8). Both *M. kudoi* and *O. scaberrima* are erect shrub and have center-positioned stigma, but they are quite different in terms of stamen morphology (dimorphic vs. isomorphic) and hypanthium indumentum



Fig. 8 Melastoma dodecandrum Lour. a Habit; b flowering branches; c inflorescence; d flower, arrow indicated the position of stigma; e longitudinal section of a flower, scale bar 10 mm; f young fruit showing the spreading hairs, scale bar 10 mm (b and d photographed by Ms. Xiaolan Wang)

Dai et al. Bot Stud (2019) 60:5 Page 9 of 11

(spreading vs. appressed hairs) (Figs. 7, 9). Geographically, both *M. kudoi* and *O. scaberrima* are restricted to Taiwan, where *M. dodecandrum* is never recorded.

Both molecular phylogenetic and morphological data suggest that *M. kudoi* is well separated from *M. intermedium*, *M. dodecandrum*, and *O. scaberrima*, and should be treated as a distinct species. Currently, only one small population of *M. kudoi* is known growing on open grassland by Lake Sui-sya. Individuals growing there were mowed from time to time, which may have impact on the survival and dispersal of this species. According to IUCN (2012), the proposed IUCN Category for this species should be CR (critically endangered).

Aside from *M. kudoi, M. suffruticosum* and *O. scaberrima* are also synonymized under *M. intermedium* (Chen and Renner 2007). *Melastoma suffruticosum* is

recorded from Hainan island. It resembles *M. intermedium* in leaf morphology, and indumentum, but differs in habit (erect vs. semicreeping). The identity of this plant remains unknown as our attempts to sample it from the field or the herbarium had both failed. The specific status of *O. scaberrima* is supported by molecular and morphological data. Its treatment will be discussed elsewhere as the generic circumscription of *Otanthera* and *Melastoma* is concerned.

Taxonomy

Melastoma kudoi Sasaki, Trans. Nat. Hist. Soc. Taiwan 21: 113, fig. 1. 1931; Liu, Ill. Native Introduced Ligneous Pl. Taiwan 1: 290, pl. 241. 1960; Yang & Liu, Taiwania 47(2): 176. 2002. Type: Taiwan, Sun Moon Lake (Lake Sui-sya), 22 Sept 1929, Kudo & Sasaki s.n. (TAI,



Fig. 9 Otanthera scaberrima (Hayata) Ohwi. a Habit; b leaves; c inflorescence; d flower, arrow indicated the position of stigma; e longitudinal section of a flower, scale bar 10 mm

Dai et al. Bot Stud (2019) 60:5 Page 10 of 11

holotype!; TAI, isotype!). *Melastoma intermedium auct. non* Dunn: Keng & Li, Fl. Taiwan 3: 855, *pl.* 841. 1977; Huang & Huang, Fl. Taiwan ed. 2. 3: 918, *pl.* 459. 1993.

Shrublets up to 50 cm tall, erect. Stems terete to slightly 4-sided, sparsely appressed strigose. Leaves opposite; petiole 2 mm; leaf blade oval, oblong ovaloblong to elliptic, $2-4 \times 1-2$ cm, papery to stiff papery, secondary veins 2 on each side of midvein, sparsely appressed strigose on both sides, but veins glabrous adaxially, base obtuse to rounded, margin more or less crenate, apex obtuse. Flowers 1-3 in terminal cymes; bracteoles lanceolate, caducous. Flowers ca. 5.5 cm in diameter, pedicels 2-3 mm, appressed strigose; calyx-tube subglobose, 9 mm long, densely strigose with spreading, basally enlarged hairs; calyx lobes 5, lanceolate, 9-12 mm long, apex acuminate, strigose; petals 5, purple, obovate, $3-3.2 \times 2.5$ cm, oblique, margin minutely ciliate; Stamens 10, unequal; filaments 8–9 mm long; anthers dimorphic; anthers of longer stamens linear-lanceolate, 6 mm long, connective decurrent to 10 mm long, 2-setose at base; anthers of shorter stamens linear-lanceolate, 5 mm long, connective slightly prolonged, 2-tuberculate at base; ovary half inferior, globose, apically strigose; style and stigma positioned in the center of the flower; style filiform, 15 mm long. Fruit a berry, globose, strigose, 7-11 mm in diameter; seeds numerous. Endemic to central

Additional specimen examined: Taiwan: Jitsugetsutan, Jan 1929, *Kudo & Sasaki s.n.* (TAIF); Nantou, Lienhuachi, 7 Jul 1929, *Kudo & Yamamoto s.n.* (TAI); Sun Moon Lake, 30 Aug 2007, *S.W. Chung 13084* (TAIF); Sun Moon Lake, 22 May 2008, *T.C. Hsu 1402* (TAIF); Sun Moon Lake, 29 June 2017, *T.C. Hsu 9315* (TAIF).

Additional file

Additional file 1: Table S1. Source of materials studied and GenBank accession numbers for nrITS, 6 nuclear genes and 2 chloroplast markers. Sequences downloaded from GenBank are indicated in bold.

Authors' contributions

RCZ and YL designed the experiments. All authors took part in the field work. CWL kept the living individuals of *M. kudoi*. YL and CWL photographed the species concerned. JHD and QJZ carried out the experiment. JHD and RCZ analyze the data. YL, JHD and RCZ wrote the first draft of the manuscript. All authors revised the manuscript. All authors revised the final manuscript.

Author details

¹ State Key Laboratory of Biocontrol and Guangdong Key Laboratory of Plant Resources, School of Life Sciences, Sun Yat-sen University, Guangzhou 510275, China. ² Herbarium of Taiwan Forestry Research Institute, No. 53, Nan-Hai Road, Taipei 100, Taiwan.

Acknowledgements

We thank Kuo-Hsiung Wang (TAI) for providing the image of the type specimen of *Melastoma kudoi*, and Ms. Xiao-Lan Wang for providing the photo of *Melastoma dodecandrum*. We are deeply grateful to the reviewers for their valuable comments on the manuscript.

Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

The datasets generated and/or analysed during the current study are available in the GenBank repository, https://www.ncbi.nlm.nih.gov/.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Not applicable.

Funding

This work was supported by the National Natural Science Foundation of China (31770214, 31670210 and 31811530297) and Science and Technology Program of Guangzhou (201707010090).

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 7 January 2019 Accepted: 16 March 2019 Published online: 28 March 2019

References

- Bandelt HJ, Forster P, Röhl A (1999) Median-joining networks for inferring intraspecific phylogenies. Mol Biol Evol 16:37–48. https://doi.org/10.1093/oxfordjournals.molbev.a026036
- Chao L-F, Chen Y-Y, Wang S-Q, Liu T, Wu W, Dai S-P, Wang F, Fan Q, Shi S-H, Zhou R-C (2014) One species or two? Multilocus analysis of nucleotide variation of *Melastoma penicillatum* and *M. sanguineum* (Melastomataceae) in Hainan, China. Biochem Syst Ecol 55:275–282. https://doi.org/10.1016/j.bse.2014.03.037
- Chen C (1984) Melastomataceae. In: Chen C (ed) Flora Reipublicae Popularis Sinicae, vol 53, no 1. Science Press, Beijing, pp 135–293
- Chen C, Renner SS (2007) Melastomataceae. In: Wu Z-Y, Raven PH (eds) Flora of China, vol 13. Science Press, Beijing, pp 360–399
- Dai S-P, Wu W, Zhang R-S, Liu T, Chen Y-Y, Shi S-H, Zhou R-C (2012) Molecular evidence for hybrid origin of *Melastoma intermedium*. Biochem Syst Ecol 41:136–141. https://doi.org/10.1016/j.bse.2011.12.010
- Doyle JJ, Doyle JL (1987) A rapid DNA isolation procedure for small amounts of fresh leaf tissue. Phytochem Bull 19:11–15
- Huang S-F, Huang T-C (1993) Melastomataceae. In: Huang T-C, Hsieh C-F, Li Z-Y, Lo H-C, Ohashi H, Shen C-F, Wang J-C, Yang K-C (eds) Flora of Taiwan, vol 3. Department of Botany, National Taiwan University, Taipei, pp 905–928
- Huang G-L, Liu Y, Wu W, Ng WL, Ning Z-L, Zhang R-J, Fan Q, Zhou R-C (2018) Multi-locus analyses indicate that *Melastoma dendrisetosum*, an endemic and endangered shrub in Hainan, is a distinct species. Syst Bot 43(1):258– 265. https://doi.org/10.1600/036364418X697120
- IUCN (2012) IUCN Red List categories and criteria: version 3.1, 2nd edn. IUCN, Gland
- Katoh K, Standley DM (2013) MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol 30:772–780. https://doi.org/10.1093/molbev/mst010
- Keng H, Li H-L (1977) Melastomataceae. In: Li H-L, Liu T-S, Huang T-C, Koyama T, DeVol CE (eds) Flora of Taiwan, vol 3. Epoch Publishing Co., Ltd., Taipei, pp 839–867
- Kimura M (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. J Mol Evol 16:111–120. https://doi.org/10.1007/BF01731581

Dai et al. Bot Stud (2019) 60:5 Page 11 of 11

- Kumar S, Stecher G, Tamura K (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol 33:1870–1874. https://doi.org/10.1093/molbev/msw054
- Liu T, Chen Y-Y, Chao L-F, Wang S-Q, Wu W, Dai S-P, Wang F, Fan Q, Zhou R-C (2014) Extensive hybridization and introgression between *Melastoma candidum* and *M. sanguineum*. PLoS ONE 9(5):e96680. https://doi.org/10.1371/journal.pone.0096680
- Meyer K (2001) Revision of the Southeast Asian genus *Melastoma*. Blumea 46(2):351–398
- Neo L, Wong KM, Tan HTW (2017) Studies in Southeast Asian *Melastoma* (Melastomataceae), 3. A new species of *Melastoma* from Peninsular Malaysia. Phytotaxa 317(4):286–291. https://doi.org/10.11646/phytotaxa.317.4.3
- Ng WL, Cai Y-C, Wu W, Zhou R-C (2017) The complete chloroplast genome sequence of *Melastoma candidum* (Melastomataceae). Mitochondrial DNA Part B Resour 2:242–243. https://doi.org/10.1080/23802359.2017.1318680
- Posada D, Crandall KA (1998) Modeltest: testing the model of DNA substitution. Bioinformatics 14:817–818. https://doi.org/10.1093/bioinformatics/14.9.817
- Renner SS, Meyer K (2001) Melastomeae come full circle: biogeographic reconstruction and molecular clock dating. Evolution 55:1315–1324. https://doi.org/10.1111/j.0014-3820.2001.tb00654.x
- Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Syst Bot 61:539–542. https://doi.org/10.1093/sysbio/sys029
- Sasaki S (1931) Miscellaneous contributions to the flora of Formosa VIII. Trans Nat Hist Soc Taiwan 21:112–118

- Stamatakis A (2014) RAXML version 8: a tool for phylogenetic analysis and post-analysis of large. Bioinformatics 30:1312–1313. https://doi.org/10.1093/bioinformatics/btu033
- Swofford DL (2002) PAUP*: phylogenetic analysis using parsimony (*and other methods), version 4.0b10. Sinauer Associates, Sunderland
- Taberlet P, Gielly L, Pautou G, Bouvet J (1991) Universal primers for amplification of three non-coding regions of chloroplast DNA. Plant Mol Biol 17:1105–1109. https://doi.org/10.1007/BF00037152
- Veranso-Libalah MC, Stone RD, Fongod AGN, Couvreur TLP, Kadereit G (2017) Phylogeny and systematics of African Melastomateae (Melastomataceae). Taxon 66(3):584–614. https://doi.org/10.12705/663.5
- White TJ, Bruns T, Lee S, Taylor JW (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (eds) PCR protocols: a guide to methods and applications. Academic Press, Inc., New York, pp 315–322. https://doi.org/10.1016/b978-0-12-372180-8.50042-1
- Wong KM (2016) The genus *Melastoma* in Borneo: including 31 new species. Natural History Publications, Kota Kinabalu
- Yang Y-P, Liu H-Y (2002) Nomenclature changes for some dicots of Taiwan. Taiwania 47(2):175–178. https://doi.org/10.6165/tai.2002.47(2).175
- Zhou Q-J, Cai Y-C, Ng WL, Wu W, Dai S-P, Wang F, Zhou R-C (2017) Molecular evidence for natural hybridization between two *Melastoma* species endemic to Hainan and their widespread congeners. Biodiversity Sci 25(6):638–646
- Zou P-S, Ng WL, Wu W, Dai S-P, Ning Z-L, Wang S-Q, Liu Y, Fan Q, Zhou R-C (2017) Similar morphologies but different origins: hybrid status of two more semi-creeping taxa of *Melastoma*. Front Plant Sci 8:673. https://doi.org/10.3389/fpls.2017.00673

Submit your manuscript to a SpringerOpen journal and benefit from:

- ► Convenient online submission
- ▶ Rigorous peer review
- ▶ Open access: articles freely available online
- ► High visibility within the field
- ► Retaining the copyright to your article

Submit your next manuscript at ▶ springeropen.com